

CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD  
LOS ANGELES REGION

MONITORING AND REPORTING PROGRAM NO. CI- 6949  
for  
MORTON INTERNATIONAL, INC.  
MORTON SALT DIVISION – LONG BEACH FACILITY  
(CA0061476)

I. Reporting Requirements

- A. Morton International, Inc. (hereinafter Morton Salt or Discharger), Morton Salt Division, Long Beach Facility (Facility) shall implement this monitoring program on the effective date of this Order. All monitoring reports shall be submitted quarterly and must be received by the Regional Board by the dates in the following schedule. All monitoring reports should be addressed to the Regional Board, Attention: Information Technology Unit. The first monitoring report under this Program is due by April 15, 2005.

Reporting Period	Report Due
January – March	April 15
April – June	July 15
July-September	October 15
October-December	January 15
Annual Summary Report	March 1

If there is no discharge during any reporting period, the report shall so state.

- B. The Discharger shall submit an annual report (for both dry and wet weather discharges from Discharge Serial Nos. 02D and 02W), containing a discussion of the previous year's effluent and receiving water monitoring data, as well as graphical and tabular summaries of the data. The data shall be submitted to the Regional Board on hard copy and on a 3 ½ " computer diskette. Submitted data must be IBM compatible, preferably using EXCEL software. This annual report is to be received by the Regional Board by March 1 of each year following the calendar year of data collection.
- C. Each monitoring report shall contain a separate section titled "Summary of Non-Compliance" which discusses the compliance record and corrective actions taken or planned that may be needed to bring the discharge into full compliance with waste discharge requirements. This section shall clearly list all non-compliance with waste discharge requirements, as well as all excursions of effluent limitations.

- D. The Discharger shall inform the Regional Board well in advance of any proposed construction activity that could potentially affect compliance with applicable requirements.

## II. Effluent Monitoring Requirements

### A. Effluent Monitoring Stations

Individual sampling designations shall be established. Each outfall shall be located where representative samples of wet scrubber effluent, salt storage pile seepage and storm water runoff (from on and off-site sources) can be obtained. The wastewater samples shall be collected from the catch basin during both dry and wet weather conditions, prior to discharging to the Long Beach Harbor.

- Discharge Serial No. 02D – Catch Basin: Samples shall be collected from the catch basin during dry weather conditions, prior to discharging through the piping from the catch basin to the final point of discharge into Long Beach Harbor.
- Discharge Serial No. 02W – Catch Basin: Samples shall be collected from the catch basin during wet weather conditions, prior to discharging through piping from the catch basin to the final point of discharge into Long Beach Harbor.

### B. Receiving Water Monitoring Station

The receiving water monitoring location shall be outside the influence of the discharge into the Long Beach Harbor; where possible, at least 50 feet from the discharge point.

- C. This Regional Board shall be notified in writing of any change in the sampling stations once established or in the methods for determining the quantities of pollutants in the individual waste streams.
- D. Pollutants shall be analyzed using the analytical methods described in 40 CFR sections 136.3, 136.4, and 136.5 (revised May 14, 1999); or, where no methods are specified for a given pollutant, by methods approved by this Regional Board or the State Board. Laboratories analyzing effluent samples and receiving water samples shall be certified by the California Department of Health Services Environmental Laboratory Accreditation Program (ELAP) or approved by the Executive Officer and must include quality assurance/quality control (QA/QC) data in their reports. A copy of

the laboratory certification shall be provided each time a new certification and/or renewal of the certification is obtained from ELAP.

The monitoring reports shall specify the analytical method used, the Method Detection Limit (MDL), and the Minimum Level (ML) for each pollutant. For the purpose of reporting compliance with numerical limitations, performance goals, and receiving water limitations, analytical data shall be reported by one of the following methods, as appropriate:

1. An actual numerical value for sample results greater than or equal to the ML; or,
2. "Detected, but Not Quantified (DNQ)" if results are greater than or equal to the laboratory's MDL but less than the ML; or,
3. "Not-Detected (ND)" for sample results less than the laboratory's MDL with the MDL indicated for the analytical method used.

Current MLs (Attachment B) are those published by the State Water Resources Control Board in the *Policy for the Implementation of Toxics Standards for Inland Surface Waters, Enclosed Bays, and Estuaries of California, March 2, 2000*.

- E. Where possible, the MLs employed for effluent analyses shall be lower than the permit limitations established for a given parameter. If the ML value is not below the effluent limitation, then the lowest ML value and its associated analytical method shall be selected for compliance purposes. At least once a year, the Discharger shall submit a list of the analytical methods employed for each test and associated laboratory QA/QC procedures.

The Regional Board, in consultation with the State Board Quality Assurance Program, shall establish a ML that is not contained in Attachment B to be included in the Discharger's permit in any of the following situations:

1. When the pollutant under consideration is not included in Attachment B;
2. When the Discharger and Regional Board agree to include in the permit a test method that is more sensitive than that specified in 40 CFR Part 136 (revised May 14, 1999);
3. When the Discharger agrees to use an ML that is lower than that listed in Attachment B;

4. When the Discharger demonstrates that the calibration standard matrix is sufficiently different from that used to establish the ML in Attachment B, and proposes an appropriate ML for their matrix; or,
  5. When the Discharger uses a method whose quantification practices are not consistent with the definition of an ML. Examples of such methods are the U.S. EPA-approved method 1613 for dioxins and furans, method 1624 for volatile organic substances, and method 1625 for semi-volatile organic substances. In such cases, the Discharger, the Regional Board, and the State Board shall agree on a lowest quantifiable limit and that limit will substitute for the ML for reporting and compliance determination purposes.
- F. Water/wastewater samples must be analyzed within allowable holding time limits as specified in 40 CFR section 136.3. All QA/QC items must be run on the same dates the samples were actually analyzed, and the results shall be reported in the Regional Board format, when it becomes available, and submitted with the laboratory reports. Proper chain of custody procedures must be followed, and a copy of the chain of custody shall be submitted with the report.
- G. All analyses shall be accompanied by the chain of custody, including but not limited to data and time of sampling, sample identification, and name of person who performed sampling, date of analysis, name of person who performed analysis, QA/QC data, method detection limits, analytical methods, copy of laboratory certification, and a perjury statement executed by the person responsible for the laboratory.
- H. Annual wet weather sampling from 02W shall be performed during the first rainfall event of the wet season (October 1 – May 31). Annual dry weather sampling for Internal Outfall 02D shall be performed during the dry season. Results of annual analyses shall be reported in the appropriate quarterly monitoring report.
- I. For parameters that both monthly average and daily maximum limits are specified and the monitoring frequency is less than four times a month, the following shall apply: If an analytical result is greater than the monthly average limit, the sampling frequency shall be increased (within 1 week of receiving the test results) to a minimum of once weekly, if possible, at equal intervals, until at least four consecutive weekly samples have been obtained, and compliance with the monthly average limit has been demonstrated. The Discharger shall provide for the approval of the Executive Officer a program to ensure future compliance with the monthly average limit.

**III. Effluent Monitoring Program**

- A. In addition to effluent monitoring of discharges to determine compliance with final effluent limitations, the Discharger must monitor the wet scrubber effluent, salt storage pile seepage and on and off-site storm water runoff for priority pollutants to determine reasonable potential. Pursuant to the California Water Code, section 13267, the Discharger is required to submit data sufficient for: (1) determining if water quality-based effluent limitations for priority pollutants are required, and (2) to calculate effluent limitations, if required. The SIP requires that the data be provided. Accordingly, the Regional Board is requiring that the Discharger conduct annual effluent monitoring of the priority pollutants listed in Section VI. The results of monitoring for reasonable potential determination shall be submitted in accordance with Section I.A of this Monitoring and Reporting Program (hereinafter *MRP*).
- B. Effluent Monitoring for Process Water (e.g., Wet Scrubber Effluent): Discharge Serial No. 02D (Dry Weather Conditions, Catch Basin)

The following shall constitute the monitoring program for continuous discharges of process water (e.g., wet scrubber effluent) through NPDES Discharge Serial No. 02D (Latitude 33° 45' 00" North and Longitude 118° 12' 30" West) in the catch basin during dry weather conditions period (April 1 – September 30) to characterize the waste stream, to collect additional information for the RPA in the future, and to determine compliance with effluent limitations:

Constituent	Units	Type of Sample	Minimum Frequency
Total Daily Waste Flow	gallons per day	Metered	Daily
Rainfall	Inches/day	Continuous	Once per discharge event
Salinity	g/L	Grab	Quarterly <sup>1</sup>
pH	s.u.	Grab	Quarterly <sup>1</sup>
Temperature	°F or °C	Grab	Quarterly <sup>1</sup>
Total Suspended Solids	mg/L	Grab	Quarterly <sup>1</sup>
Oil and Grease	mg/L	Grab	Quarterly <sup>1</sup>
Settleable Solids	ml/L	Grab	Quarterly <sup>1</sup>
Turbidity	NTU	Grab	Quarterly <sup>1</sup>
BOD <sub>5</sub> @ 20 °C	mg/L	Grab	Annually <sup>2</sup>
2,3,7,8-TCDD (dioxin) and	µg/L	Grab	Once <sup>3</sup>

Constituent	Units	Type of Sample	Minimum Frequency
congeners			
CTR Priority Pollutants (see pages T-14 - 16 of the MRP)	µg/L	Grab	Annually <sup>2</sup>

1. Sampling shall be conducted for two quarters (April 1 – June 30, July 1 – September 30) per year during the dry weather condition period.
2. Sampling shall be conducted once per year during the dry weather condition period (April 1 – September 30).
3. Sampling shall be conducted once during the dry weather condition period (April 1, 2005 – September 30, 2005).

This monitoring schedule is effective upon adoption of the Order by the Regional Board.

- C. Effluent Monitoring for Process Water (e.g., Wet Scrubber Effluent), Salt Storage Pile Seepage and On-Site/Off-Site Storm Water Runoff: Discharge Serial No. 02W (Wet Weather Conditions, Catch Basin)

The following shall constitute the monitoring program for continuous discharges of process water (e.g., wet scrubber effluent), and storm water runoff (e.g., salt storage pile seepage and on-site storm water runoff, and off-site storm water runoff from an adjacent property) through NPDES Discharge Serial No. 02W (Latitude 33° 45' 00" North and Longitude 118° 12' 30" West) in the catch basin during wet weather conditions to characterize the waste stream, collect additional information for the RPA in the future, and to determine compliance with the final effluent limitations:

Constituent	Units	Type of Sample	Minimum Frequency
Total Daily Waste Flow	gallons per day	Metered	Daily
Rainfall	Inches/day	Continuous	Once per discharge event
Salinity	g/L	Grab	Quarterly <sup>1</sup>
PH	s.u.	Grab	Quarterly <sup>1</sup>
Temperature	°F or °C	Grab	Quarterly <sup>1</sup>
Total Suspended Solids	mg/L,	Grab	Quarterly <sup>1</sup>
Oil and Grease	mg/L,	Grab	Quarterly <sup>1</sup>
Settleable Solids	ml/L	Grab	Quarterly <sup>1</sup>
Turbidity	NTU	Grab	Quarterly <sup>1</sup>
BOD <sub>5</sub> @ 20 °C	mg/L	Grab	Annually <sup>2</sup>

Constituent	Units	Type of Sample	Minimum Frequency
2,3,7,8-TCDD (dioxin) and congeners	µg/L	Grab	Once <sup>3</sup>
CTR Priority Pollutants (see pages T-14 – 17 of the MRP)	µg/L	Grab	Annually <sup>2</sup>
Acute Toxicity	% Survival	Grab	Annually <sup>2</sup>

1. Sampling shall be conducted for two quarters (October 1 – December 31, January 1 – March 31) per year during the wet weather condition period.
2. Sampling shall be conducted once per year during the wet weather condition period (October 1 – March 31).
3. Sampling shall be conducted once during the wet weather condition period (October 1, 2005 – March 31, 2006).

This monitoring schedule is effective upon adoption of the Order by the Regional Board.

#### IV. Toxicity Monitoring Requirements

##### A. Acute Toxicity Effluent Monitoring Program

1. The Discharger shall conduct acute toxicity tests on effluent grab samples by methods specified in 40 CFR Part 136 which cites U.S. EPA's *Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms*, Fifth Edition, October 2002, U.S. EPA, Office of Water, Washington D.C. (EPA/821-R-02-012) or a more recent edition to ensure compliance in 100 % effluent.
2. The fathead minnow, *Pimephales promelas*, shall be used as the test species for fresh water discharges and the topsmelt, *Atherinops affinis*, shall be used as the test species for brackish effluent. The method for topsmelt is found in U.S. EPA's *Short-term Method for Estimating the Chronic Toxicity of Effluents and Receiving Waters to West Coast Marine and Estuarine Organisms*, Third Edition, October 2002 (EPA/821-R-02-014).
3. In lieu of conducting the standard acute toxicity testing with the fathead minnow, the Discharger may elect to report the results or endpoint from the first 48 hours of the chronic toxicity test as the results of the acute toxicity test.

4. Effluent samples shall be collected after all treatment processes and before discharge to the receiving water.

B. Quality Assurance

1. Concurrent testing with a reference toxicant shall be conducted. Reference toxicant tests shall be conducted using the same test conditions as the effluent toxicity tests (e.g., same test duration, etc).
2. If either the reference toxicant test or effluent test does not meet all test acceptability criteria (TAC) as specified in the test methods manuals (EPA/821-R-02-013 and EPA/821-R-02-014), then the Discharger must re-sample and re-test at the earliest time possible.
3. Control and dilution water should be receiving water or laboratory water, as appropriate, as described in the manual. If the dilution water used is different from the culture water, a second control using culture water shall be used.

C. Accelerated Monitoring

1. If toxicity exceeds the limitations (as defined in Order No. R4-2004-0174, Sections I.B.3.a.i.), then the Discharger shall immediately implement accelerated testing as specified in Sections I.B.3.a.ii. The Discharger shall ensure that they receive results of a failing acute toxicity test within 24 hours of the close of the test and the additional tests shall begin within three business days of the receipt of the result. If the accelerated testing shows consistent toxicity, the Discharger shall immediately implement the Initial Investigation of the Toxicity Reduction Evaluation (TRE) Workplan.
2. If implementation of the initial investigation TRE Workplan indicates the source of toxicity (e.g., a temporary plant upset, etc.), then the Discharger may discontinue the Toxicity Identification Evaluation (TIE).
3. The first step in the initial Investigation TRE Workplan for downstream receiving water toxicity can be a toxicity test protocol designed to determine if the effluent from Discharge Serial No. 001 causes or contributes to the measured downstream acute toxicity. If this first step TRE testing shows that the Discharge Serial No. 001 effluent does not cause or contribute to downstream acute toxicity, using U.S. EPA's *Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms*, Fifth Edition, October 2002, U.S. EPA, Office of Water,



Washington D.C. (EPA/821-R-02-012), then a report on this testing shall be submitted to the Board and the TRE will be considered to be completed. Routine testing in accordance with MRP No.6949 shall be continued thereafter.

D. Steps in TRE and TIE procedures:

1. Following a TRE trigger, the Discharger shall initiate a TRE in accordance with the facility's initial investigation TRE workplan. The Discharger shall use EPA manuals EPA/600/2-88/070 (industrial) or EPA/833B-99/002 (municipal) as guidance or current versions. At a minimum, the TRE workplan must contain the provision in Attachment C. The Discharger shall expeditiously develop a more detailed TRE workplan for submittal to the Executive Officer within 30 days of the trigger, which will include, but not be limited to:
  - a. Further actions to investigate and identify the cause of toxicity;
  - b. Actions the Discharger will take to mitigate the impact of the discharge and prevent the recurrence of toxicity;
  - c. Standards the Discharger will apply to consider the TRE complete and to return to normal sampling frequency; and,
  - d. A schedule for these actions.
2. The following is a stepwise approach in conducting the TRE:
  - a. Step 1 - Basic data collection. Data collected for the accelerated monitoring requirements may be used to conduct the TRE:
  - b. Step 2 - Evaluates optimization of the treatment system operation, facility housekeeping, and the selection and use of in-plant process chemicals;
  - c. If Steps 1 and 2 are unsuccessful, Step 3 implements a TIE and employment of all reasonable efforts and using currently available TIE methodologies. The objective of the TIE is to identify the substance or combination of substances causing the observed toxicity;
  - d. Assuming successful identification or characterization of the toxicant(s), Step 4 evaluates final effluent treatment options;
  - e. Step 5 evaluates in-plant treatment options; and,

- f. Step 6 consists of confirmation once a toxicity control method has been implemented.

Many recommended TRE elements parallel source control, pollution prevention, and storm water control program best management practices (BMPs). To prevent duplication of efforts, evidence of implementation of these control measures may be sufficient to comply with TRE requirements. By requiring the first steps of a TRE to be accelerated testing and review of the facility's TRE workplan, a TRE may be ended in its early stages. All reasonable steps shall be taken to reduce toxicity to the required level. The TRE may be ended at any stage if monitoring indicates there is no longer toxicity (or six consecutive chronic toxicity results are less than or equal to 1.0 TU<sub>c</sub>).

3. The Discharger may initiate a TIE as part of the TRE process to identify the cause(s) of toxicity. The Discharger shall use the EPA acute and chronic manuals, EPA/600/6-91/005F (Phase I)/EPA/600/R-96-054 (for marine), EPA/600/R-92/080 (Phase II), and EPA-600/R-92/081 (Phase III) as guidance.
4. If a TRE/TIE is initiated prior to completion of the accelerated testing schedule required by Part I.B.3.a.ii of this permit, then the accelerated testing schedule may be terminated, or used as necessary in performing the TRE/TIE, as determined by the Executive Officer.
5. Toxicity tests conducted as part of a TRE/TIE may also be used for compliance, if appropriate.
6. The Board recognizes that toxicity may be episodic and identification of causes of and reduction of sources of toxicity may not be successful in all cases. Consideration of enforcement action by the Board will be based in part on the Discharger's actions and efforts to identify and control or reduce sources of consistent toxicity.

#### E. Reporting

1. The Discharger shall submit a full report of the toxicity test results, including any accelerated testing conducted during the month as required by this permit. Test results shall be reported as % survival with the discharge monitoring reports (DMR) for the month in which the test is conducted.
2. If an initial investigation indicates the source of toxicity and accelerated testing is unnecessary, then those results also shall be submitted with the DMR for the

period in which the investigation occurred.

- a. The full report shall be submitted on or before the end of the month in which the DMR is submitted.
  - b. The full report shall consist of (1) the results; (2) the dates of sample collection and initiation of each toxicity test; (3) the acute toxicity average limit or chronic toxicity limit or trigger.
3. Test results for toxicity tests also shall be reported according to the appropriate manual chapter on Report Preparation and shall be attached to the DMR. Routine reporting shall include, at a minimum, as applicable, for each test:
- a. Sample date(s);
  - b. Test initiation date;
  - c. Test species;
  - d. End point values for each dilution (e.g., number of young, growth rate, percent survival);
  - e. NOEC value(s) in percent effluent;
  - f.  $IC_{15}$ ,  $IC_{25}$ ,  $IC_{40}$  and  $IC_{50}$  values in percent effluent;
  - g.  $TU_c$  values  $\left( TU_c = \frac{100}{NOEC} \right)$ ;
  - h. Mean percent mortality ( $\pm$ standard deviation) after 96 hours in 100% effluent (if applicable);
  - i. NOEC and LOEC values for reference toxicant test(s);
  - j.  $C_{25}$  value for reference toxicant test(s);
  - k. Any applicable charts; and
  - l. Available water quality measurements for each test (e.g., pH, D.O., temperature, conductivity, hardness, salinity, ammonia).
4. The Discharger shall provide a compliance summary, which includes a summary table of toxicity data from all samples collected during that year.

The Discharger shall notify by telephone or electronically, this Regional Board of any toxicity exceedance of the limit or trigger within 24 hours of receipt of the results followed by a written report within 14 calendar days of receipt of the results. The verbal or electronic notification shall include the exceedance and the plan the Discharger has taken or will take to investigate and correct the cause(s) of toxicity. It may also include a status report on any actions required by the permit, with a schedule for actions not yet completed. If no actions have been taken, the reasons shall be given.

**V. Receiving Water Monitoring**

The Discharger is required to monitor the receiving water for the CTR priority pollutants to determine reasonable potential. Pursuant to the California Water Code, section 13267, the Discharger is required to submit data sufficient for: (1) determining if WQBELs for priority pollutants are required, and (2) to calculate effluent limitations, if required. The SIP (March 2, 2000) requires the Discharger to provide the data. Therefore, the Discharger shall conduct annual monitoring of the receiving water for all CTR priority pollutants, pH, and hardness for first two years after adoption of the permit. The results of monitoring for reasonable potential determination shall be submitted in accordance with Section I.A of the MRP. Receiving water sampling shall be conducted at the same time as effluent sampling of Discharge Serial No. 02W. The receiving water monitoring location shall be outside the influence of the discharge into the receiving water (e.g., Long Beach Harbor), where possible, at least 50 feet from the discharge point into Long Beach Harbor.

The required monitoring frequency and type of sample of the receiving water for toxic pollutants are listed below:

Constituent	Units	Type of Sample	Monitoring Frequency
pH	standard units	Grab	Annually <sup>1</sup>
Hardness (as CaCO <sub>3</sub> )	mg/L	Grab	Annually <sup>1</sup>
2,3,7,8-TCDD (dioxin) and congeners	µg/L	Grab	Once <sup>2</sup>
CTR - Priority Pollutants (see pages T-13 – 16)	µg/L	Grab	Annually <sup>1</sup>

1. The Discharger shall collect and analyze receiving water samples annually for the first two years after adoption of the permit.
2. Sampling shall be conducted once during the wet weather condition period (October 1, 2005 – March 31, 2006).

To fulfill the receiving water priority pollutant monitoring requirement, the Discharger may elect to enter into a collaborative receiving water sampling program with other Dischargers if the point of discharge into the receiving water is shared by the Dischargers. By entering into a collaborative sampling program, the Discharger is still required to submit receiving water data for pH, hardness, and all CTR priority pollutants to the Regional Board.

**VI. Effluent and Receiving Water Monitoring for Reasonable Potential Determination**

- A. As described in Sections III and V of this *MRP*, the Discharger is required to monitor both the effluent and receiving water for priority pollutants listed in the table below, annually, for the life of the permit, to determine reasonable potential.
- B. Monitoring for reasonable potential determination shall occur at the following locations, as described previously:
- Discharge Serial No. 02D;
  - Discharge Serial No. 02W; and
  - Receiving Water.

The following provides a listing of CTR priority pollutants, required monitoring frequency and type of sample for monitoring the effluent and receiving water for reasonable potential determination:

Pollutant	Units	Type of Sample	Monitoring Frequency
Antimony	µg/L	Grab	Annually
Arsenic <sup>1</sup>	µg/L	Grab	Annually
Beryllium	µg/L	Grab	Annually
Chromium (III) <sup>1</sup>	µg/L	Grab	Annually
Chromium (VI) <sup>1</sup>	µg/L	Grab	Annually
Copper <sup>1</sup>	µg/L	Grab	Annually
Lead <sup>1</sup>	µg/L	Grab	Annually
Mercury	µg/L	Grab	Annually
Nickel <sup>1</sup>	µg/L	Grab	Annually
Selenium	µg/L	Grab	Annually
Silver <sup>1</sup>	µg/L	Grab	Annually
Thallium	µg/L	Grab	Annually
Zinc <sup>1</sup>	µg/L	Grab	Annually
Cyanide	µg/L	Grab	Annually
Asbestos	Fibers/L	Grab	Annually
Acrolein	µg/L	Grab	Annually
Acrylonitrile	µg/L	Grab	Annually
Benzene	µg/L	Grab	Annually
Bromoform	µg/L	Grab	Annually
Carbon tetrachloride	µg/L	Grab	Annually
Chlorobenzene	µg/L	Grab	Annually
Chlorodibromomethane	µg/L	Grab	Annually

Pollutant	Units	Type of Sample	Monitoring Frequency
Chloroethane	µg/L	Grab	Annually
2-Chloroethylvinyl ether	µg/L	Grab	Annually
Chloroform	µg/L	Grab	Annually
Dichlorobromomethane	µg/L	Grab	Annually
1,1-Dichloroethane	µg/L	Grab	Annually
1,2-Dichloroethane	µg/L	Grab	Annually
1,1-Dichloroethylene	µg/L	Grab	Annually
1,2-Dichloropropane	µg/L	Grab	Annually
1,3-Dichloropropylene	µg/L	Grab	Annually
Ethylbenzene	µg/L	Grab	Annually
Methyl bromide	µg/L	Grab	Annually
Methyl chloride	µg/L	Grab	Annually
Methylene chloride	µg/L	Grab	Annually
1,1,2,2-Tetrachloroethane	µg/L	Grab	Annually
Tetrachloroethylene	µg/L	Grab	Annually
Toluene	µg/L	Grab	Annually
1,2-Trans-dichloroethylene	µg/L	Grab	Annually
1,1,1-Trichloroethane	µg/L	Grab	Annually
1,1,2-Trichloroethane	µg/L	Grab	Annually
Trichloroethylene	µg/L	Grab	Annually
Vinyl Chloride	µg/L	Grab	Annually
2-Chlorophenol	µg/L	Grab	Annually
2,4-Dichlorophenol	µg/L	Grab	Annually
2,4-Dimethylphenol	µg/L	Grab	Annually
2-Methyl-4,6-Dinitrophenol	µg/L	Grab	Annually
2,4-Dinitrophenol	µg/L	Grab	Annually
2-Nitrophenol	µg/L	Grab	Annually
4-Nitrophenol	µg/L	Grab	Annually
3-Methyl-4-Chlorophenol	µg/L	Grab	Annually
Pentachlorophenol	µg/L	Grab	Annually
Phenol	µg/L	Grab	Annually
2,4,6-Trichlorophenol	µg/L	Grab	Annually
Acenaphthene	µg/L	Grab	Annually
Acenaphthylene	µg/L	Grab	Annually
Anthracene	µg/L	Grab	Annually
Benzidine	µg/L	Grab	Annually
Benzo(a)Anthracene	µg/L	Grab	Annually
Benzo(a)Pyrene	µg/L	Grab	Annually

Pollutant	Units	Type of Sample	Monitoring Frequency
Benzo(b)Fluoranthene	µg/L	Grab	Annually
Benzo (g,h,i)Perylene	µg/L	Grab	Annually
Benzo(k)Fluoranthene	µg/L	Grab	Annually
Bis (2-Chloroethoxy)Methane	µg/L	Grab	Annually
Bis(2-Chloroethyl)Ether	µg/L	Grab	Annually
Bis (2-Chloroisopropyl)Ether	µg/L	Grab	Annually
4-Bromophenyl Phenyl Ether	µg/L	Grab	Annually
Butylbenzyl Phthalate	µg/L	Grab	Annually
2-Chloronaphthalene	µg/L	Grab	Annually
4-Chlorophenyl Phenyl Ether	µg/L	Grab	Annually
Chrysene	µg/L	Grab	Annually
Dibenzo(a,h)Anthracene	µg/L	Grab	Annually
1,2-Dichlorobenzene	µg/L	Grab	Annually
1,3-Dichlorobenzene	µg/L	Grab	Annually
1,4-Dichlorobenzene	µg/L	Grab	Annually
3,3'-Dichlorobenzidine	µg/L	Grab	Annually
Diethyl Phthalate	µg/L	Grab	Annually
Dimethyl Phthalate	µg/L	Grab	Annually
Di-n-Butyl Phthalate	µg/L	Grab	Annually
2,4-Dinitrotoluene	µg/L	Grab	Annually
2,6-Dinitrotoluene	µg/L	Grab	Annually
Di-n-Octyl Phthalate	µg/L	Grab	Annually
1,2-Diphenylhydrazine	µg/L	Grab	Annually
Fluoranthene	µg/L	Grab	Annually
Fluorene	µg/L	Grab	Annually
Hexachlorobenzene	µg/L	Grab	Annually
Hexachlorobutadiene	µg/L	Grab	Annually
Hexachlorocyclopentadiene	µg/L	Grab	Annually
Hexachloroethane	µg/L	Grab	Annually
Indeno(1,2,3-cd)Pyrene	µg/L	Grab	Annually
Isophorone	µg/L	Grab	Annually
Naphthalene	µg/L	Grab	Annually
Nitrobenzene	µg/L	Grab	Annually
N-Nitrosodimethylamine	µg/L	Grab	Annually
N-Nitrosodi-n-Propylamine	µg/L	Grab	Annually
N-Nitrosodiphenylamine	µg/L	Grab	Annually
Phenanthrene	µg/L	Grab	Annually
Pyrene	µg/L	Grab	Annually

Pollutant	Units	Type of Sample	Monitoring Frequency
1,2,4-Trichlorobenzene	µg/L	Grab	Annually
Aldrin	µg/L	Grab	Annually
Alpha-BHC	µg/L	Grab	Annually
beta-BHC	µg/L	Grab	Annually
Gamma-BHC	µg/L	Grab	Annually
Delta-BHC	µg/L	Grab	Annually
Chlordane	µg/L	Grab	Annually
4,4'-DDT	µg/L	Grab	Annually
4,4'-DDE	µg/L	Grab	Annually
4,4'-DDD	µg/L	Grab	Annually
Dieldrin	µg/L	Grab	Annually
Alpha-Endosulfan	µg/L	Grab	Annually
beta-Endosulfan	µg/L	Grab	Annually
Endosulfan Sulfate	µg/L	Grab	Annually
Endrin	µg/L	Grab	Annually
Endrin Aldehyde	µg/L	Grab	Annually
Heptachlor	µg/L	Grab	Annually
Heptachlor Epoxide	µg/L	Grab	Annually
PCBs – Sum <sup>2</sup>	µg/L	Grab	Annually
Toxaphene	µg/L	Grab	Annually

<sup>1</sup> Measured as total recoverable.

<sup>2</sup> The sum of PCB Arochlors 1016, 1221, 1232, 1242, 1248, 1254, and 1260.

- C. In accordance with Section 3 of the SIP, the Discharger is also required to conduct effluent and receiving water monitoring for the presence of the 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD or Dioxin) congeners. The monitoring shall be a grab sample conducted once during the first year. The Discharger is required to monitor for 2,3,7,8-TCDD and the 16 congeners listed in the table below. The Discharger is required to calculate Toxic Equivalence (TEQ) for each congener by multiplying its analytical concentration by the appropriate Toxicity Equivalence Factors (TEF) provided below.

Congeners	TEF
2,3,7,8-tetra CDD	1.0
1,2,3,7,8-penta CDD	1.0
1,2,3,4,7,8-hexa CDD	0.1
1,2,3,6,7,8-hexa CDD	0.1
1,2,3,7,8,9-hexa CDD	0.1
1,2,3,4,6,7,8-hepta CDD	0.01



<b>Congeners</b>	<b>TEF</b>
Octa CDD	0.0001
2,3,7,8-tetra CDF	0.1
1,2,3,7,8-penta CDF	0.05
2,3,4,7,8-penta CDF	0.5
1,2,3,4,7,8-hexa CDF	0.1
1,2,3,6,7,8-hexa CDF	0.1
1,2,3,7,8,9-hexa CDF	0.1
2,3,4,6,7,8-hexa CDF	0.1
1,2,3,4,6,7,8-hepta CDF	0.01
1,2,3,4,7,8,9-hepta CDF	0.01
Octa CDF	0.0001

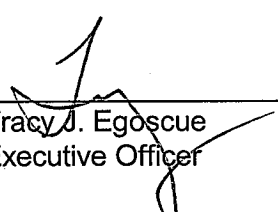
- D. The reports for this required monitoring must be submitted separately from the self-monitoring reports, but in accordance with the quarterly reporting schedule provided in Section I.A. The reports shall reference "Monitoring Results for CTR Priority Pollutants Reasonable Potential Determination".
- E. SWRCB-approved laboratory methods and the corresponding MLs for the examination of each priority pollutant are listed in Attachment B-1. Reporting requirements for the data to be submitted are listed in Attachment C. The Regional Board recommends that an analytical method be selected from Attachment B which is capable of achieving the lowest ML for each pollutant as listed on Attachment B-1. ML is necessary for determining compliance for a priority pollutant when an effluent limit is below the MDL.
- F. The laboratory analytical data shall include applicable MLs, MDL, quality assurance/quality control data, and shall comply with the reporting requirements contained in the Attachments B-1 & C.
- G. Forward all interim monitoring data/reports to The Regional Board, Attn: Industrial Permitting Unit, and please include a reference to "Compliance File No. CI-6949, NPDES No. CA0061476."

**VII. Storm Water Monitoring Requirements**

**Rainfall Monitoring**

The Discharger shall measure and record the rainfall on each day of the month. This information shall be included in the monitoring report for that month.

Ordered by:

  
\_\_\_\_\_  
Tracy J. Egoscue  
Executive Officer

Date: February 5, 2009

SWRCB Minimum Levels in ppb ( $\mu\text{g/L}$ )

The Minimum Levels (MLs) in this appendix are for use in reporting and compliance determination purposes in accordance with section 2.4 of the State Implementation Policy. These MLs were derived from data for priority pollutants provided by State certified analytical laboratories in 1997 and 1998. These MLs shall be used until new values are adopted by the SWRCB and become effective. The following tables (Tables 2a - 2d) present MLs for four major chemical groupings: volatile substances, semi-volatile substances, inorganics, and pesticides and PCBs.

Table 2a - VOLATILE SUBSTANCES*	GC	GCMS
1,1 Dichloroethane	0.5	1
1,1 Dichloroethylene	0.5	2
1,1,1 Trichloroethane	0.5	2
1,1,2 Trichloroethane	0.5	2
1,1,2,2 Tetrachloroethane	0.5	1
1,2 Dichlorobenzene (volatile)	0.5	2
1,2 Dichloroethane	0.5	2
1,2 Dichloropropane	0.5	1
1,3 Dichlorobenzene (volatile)	0.5	2
1,3 Dichloropropene (volatile)	0.5	2
1,4 Dichlorobenzene (volatile)	0.5	2
Acrolein	2.0	5
Acrylonitrile	2.0	2
Benzene	0.5	2
Bromoform	0.5	2
Methyl Bromide	1.0	2
Carbon Tetrachloride	0.5	2
Chlorobenzene	0.5	2
Chlorodibromo-methane	0.5	2
Chloroethane	0.5	2
Chloroform	0.5	2
Chloromethane	0.5	2
Dichlorobromo-methane	0.5	2
Dichloromethane	0.5	2
Ethylbenzene	0.5	2
Tetrachloroethylene	0.5	2
Toluene	0.5	2
Trans-1,2 Dichloroethylene	0.5	1
Trichloroethene	0.5	2
Vinyl Chloride	0.5	2

\*The normal method-specific factor for these substances is 1; therefore, the lowest standard concentration in the calibration curve is equal to the above ML value for each substance.

## Attachment B - continued

Table 2b - SEMI-VOLATILE SUBSTANCES*	GC	GCMS	LC	COLOR
Benzo (a) Anthracene	10	5		
1,2 Dichlorobenzene (semivolatile)	2	2		
1,2 Diphenylhydrazine		1		
1,2,4 Trichlorobenzene	1	5		
1,3 Dichlorobenzene (semivolatile)	2	1		
1,4 Dichlorobenzene (semivolatile)	2	1		
2 Chlorophenol	2	5		
2,4 Dichlorophenol	1	5		
2,4 Dimethylphenol	1	2		
2,4 Dinitrophenol	5	5		
2,4 Dinitrotoluene	10	5		
2,4,6 Trichlorophenol	10	10		
2,6 Dinitrotoluene		5		
2- Nitrophenol		10		
2-Chloroethyl vinyl ether	1	1		
2-Chloronaphthalene		10		
3,3' Dichlorobenzidine		5		
Benzo (b) Fluoranthene		10	10	
3-Methyl-Chlorophenol	5	1		
4,6 Dinitro-2-methylphenol	10	5		
4- Nitrophenol	5	10		
4-Bromophenyl phenyl ether	10	5		
4-Chlorophenyl phenyl ether		5		
Acenaphthene	1	1	0.5	
Acenaphthylene		10	0.2	
Anthracene		10	2	
Benzidine		5		
Benzo(a) pyrene		10	2	
Benzo(g,h,i)perylene		5	0.1	
Benzo(k)fluoranthene		10	2	
bis 2-(1-Chloroethoxyl) methane		5		
bis(2-chloroethyl) ether	10	1		
bis(2-Chloroisopropyl) ether	10	2		
bis(2-Ethylhexyl) phthalate	10	5		
Butyl benzyl phthalate	10	10		
Chrysene		10	5	
di-n-Butyl phthalate		10		
di-n-Octyl phthalate		10		
Dibenzo(a,h)-anthracene		10	0.1	
Diethyl phthalate	10	2		
Dimethyl phthalate	10	2		
Fluoranthene	10	1	0.05	
Fluorene		10	0.1	

## Attachment B - continued

Table 2b - SEMI-VOLATILE SUBSTANCES*	GC	GCMS	LC	COLOR
Hexachloro-cyclopentadiene	5	5		
Hexachlorobenzene	5	1		
Hexachlorobutadiene	5	1		
Hexachloroethane	5	1		
Indeno(1,2,3,cd)-pyrene		10	0.05	
Isophorone	10	1		
N-Nitroso diphenyl amine	10	1		
N-Nitroso-dimethyl amine	10	5		
N-Nitroso -di n-propyl amine	10	5		
Naphthalene	10	1	0.2	
Nitrobenzene	10	1		
Pentachlorophenol	1	5		
Phenanthrene		5	0.05	
Phenol **	1	1		50
Pyrene		10	0.05	

\* With the exception of phenol by colorimetric technique, the normal method-specific factor for these substances is 1,000; therefore, the lowest standard concentration in the calibration curve is equal to the above ML value for each substance multiplied by 1,000.

\*\* Phenol by colorimetric technique has a factor of 1.

Table 2c - INORGANICS*	FAA	GFAA	ICP	ICPMS	SPGFAA	HYDRIDE	CVAA	COLOR	DCP
Antimony	10	5	50	0.5	5	0.5			1,000
Arsenic		2	10	2	2	1		20	1,000
Beryllium	20	0.5	2	0.5	1				1,000
Cadmium	10	0.5	10	0.25	0.5				1,000
Chromium (total)	50	2	10	0.5	1				1,000
Chromium VI	5							10	
Copper	25	5	10	0.5	2				1,000
Cyanide								5	
Lead	20	5	5	0.5	2				10,000
Mercury				0.5			0.2		
Nickel	50	5	20	1	5				1,000
Selenium		5	10	2	5	1			1,000
Silver	10	1	10	0.25	2				1,000
Thallium	10	2	10	1	5				1,000
Zinc	20		20	1	10				1,000

\* The normal method-specific factor for these substances is 1; therefore, the lowest standard concentration in the calibration curve is equal to the above ML value for each substance.

Table 2d. - PESTICIDES - PCBs*	GC
4,4'-DDD	0.05
4,4'-DDE	0.05
4,4'-DDT	0.01
a-Endosulfan	0.02
alpha-BHC	0.01
Aldrin	0.005
b-Endosulfan	0.01
Beta-BHC	0.005
Chlordane	0.1
Delta-BHC	0.005
Dieldrin	0.01
Endosulfan Sulfate	0.05
Endrin	0.01
Endrin Aldehyde	0.01
Heptachlor	0.01
Heptachlor Epoxide	0.01
Gamma-BHC (Lindane)	0.02
PCB 1016	0.5
PCB 1221	0.5
PCB 1232	0.5
PCB 1242	0.5
PCB 1248	0.5
PCB 1254	0.5
PCB 1260	0.5
Toxaphene	0.5

\* The normal method-specific factor for these substances is 100; therefore, the lowest standard concentration in the calibration curve is equal to the above ML value for each substance multiplied by 100.

#### Techniques:

GC - Gas Chromatography

GCMS - Gas Chromatography/Mass Spectrometry

HRGCMS - High Resolution Gas Chromatography/Mass Spectrometry (i.e., EPA 1613, 1624, or 1625)

LC - High Pressure Liquid Chromatography

FAA - Flame Atomic Absorption

GFAA - Graphite Furnace Atomic Absorption

HYDRIDE - Gaseous Hydride Atomic Absorption

CVAA - Cold Vapor Atomic Absorption

ICP - Inductively Coupled Plasma

ICPMS - Inductively Coupled Plasma/Mass Spectrometry

SPGFAA - Stabilized Platform Graphite Furnace Atomic Absorption (i.e., EPA 200.9)

DCP - Direct Current Plasma

COLOR - Colorimetric

**ATTACHMENT B-1**

**ANALYTICAL METHODS FOR INTERIM MONITORING**

#	Compound	Pollutant ID	Toxic Pollutants	EPA Analytical Method
			Metals & Miscellaneous	
1		1097	Antimony (Sb)	200.7, 200.8, 204.1, 204.2, 6010B, 6020, 7040, 7041
2		1000	Arsenic (As)	200.7, 200.8, 200.9, 206.2, 206.3, 206.4, 206.5, 6010B, 6020, 7060A, 7061A
3		1012	Beryllium (Be)	200.7, 200.8, 200.9, 210.1, 210.2, 6010B, 6020, 7090, 7091
4		1027	Cadmium (Cd)	200.7, 200.8, 200.9, 213.1, 213.2, 6010B, 6020, 7130, 7131A
5a		1032	Chromium (Total)	200.7, 200.8, 200.9, 218.1, 218.2, 218.3, 6010B, 6020, 7190, 7191
5b		1033	Chromium-(Cr-VI)	218.4, 7196A, 218.6, 719.9
6		1119	Copper (Cu)	200.7, 200.8, 200.9, 220.1, 220.2, 6010B, 6020, 7210, 7211
		720	Cyanide (CN)	335.2, 335.3, 9010B, 9012A
8		1051	Lead (Pb)	200.8, 200.9, 239.1, 239.2, 6010B, 6020, 7420, 7421
9		71900	Mercury (Hg)	245.1, 245.2, 200.8, 7470A, 7471A
10		1067	Nickel (Ni)	200.7, 200.8, 200.9, 249.1, 249.2, 6010B, 6020, 7520, 7521
11		1147	Selenium (Se)	200.7, 200.8, 200.9, 270.2, 6010B, 6020, 7740, 7741A
12		1077	Silver (Ag)	200.7, 200.8, 200.9, 272.1, 272.2, 6010B, 6020, 7760A, 7761
13		1059	Thallium (Tl)	200.7, 200.8, 200.9, 279.1, 279.2, 6010B, 6020, 7840, 7841
14		1092	Zinc (Zn)	200.7, 200.8, 289.1, 289.2, 6010B, 6020, 7950, 7951
15		948	Asbestos	100.1, 100.2
16 <sup>2</sup>		82698	TCDD Equivalent Toxicity Equivalent Factors (TEFs) for 2,3,7,8-TCDD Equivalents Congener	8280A, 8290
			TEF	
			2,3,7,8-TetraCDD	1
			1,2,3,7,8-PentaCDD	1.0
			1,2,3,4,7,8-HexaCDD	0.1
			1,2,3,6,7,8-HexaCDD	0.1
			1,2,3,7,8,9-HexaCDD	0.1
			1,2,3,4,6,7,8-HeptaCDD	0.01
			OctaCDD	0.0001
			2,3,7,8-TetraCDF	0.1

<sup>1</sup> Analytical Method selected must be capable of achieving an ML that is lower than the lowest criterion for the pollutant, as shown on Attachment B.

<sup>2</sup> You shall report for each congener the analytical results of the effluent monitoring, including the quantifiable limit and the MDL, and the measured or estimated concentration. In addition you shall multiply each measured or estimated congener concentration by its respective TEF value above and report the sum of these values.

**ATTACHMENT B-1**

# Compound	Pollutant ID	Toxic Pollutants	EPA Analytical Method <sup>1</sup>
		1,2,3,7,8-PentaCDF	0.05
		2,3,4,7,8-PentaCDF	0.5
		1,2,3,4,7,8-HexaCDF	0.1
		1,2,3,6,7,8-HexaCDF	0.1
		1,2,3,7,8,9-HexaCDF	0.1
		2,3,4,6,7,8-HexaCDF	0.1
		1,2,3,4,6,7,8-HeptaCDF	0.01
		1,2,3,4,7,8,9-HeptaCDF	0.01
		OctaCDF	0.0001
		<b>Volatile Pollutants</b>	
17	34210	Acrolein	603, 8030A, 8260B
18	34215	Acrylonitrile	603, 8031, 8260B
19	34030	Benzene	602, 624, 8021B, 8260B
20	32104	Bromoform	601, 624, 8021B, 8260B
21	32102	Carbon Tetrachloride	601, 624, 8021B, 8260B
22	34301	Chlorobenzene	601, 602, 624, 8021B, 8260B
23	34306	Chlorodibromomethane	601, 624, 8021B, 8260B
24	85811	Chloroethane	601, 624, 8021B, 8260B
25	34576	2-Chloroethylvinyl Ether	601, 624, 8021B, 8260B
26	32106	Chloroform	601, 624, 8021B, 8260B
27	32101	Dichlorobromomethane	601, 624, 8021B, 8260B
28	34496	1,1-Dichloroethane	601, 624, 8021B, 8260B
29	32103	1,2-Dichloroethane	601, 624, 8021B, 8260B
30	34501	1,1-Dichloroethylene	601, 624, 8021B, 8260B
31	34541	1,2-Dichloropropane	601, 624, 8021B, 8260B
32	34561	1,3-Dichloropropylene	601, 624, 8021B, 8260B
33	78113	Ethylbenzene	602, 624, 8021B, 8260B
34	34413	Methyl Bromide	601, 624, 8021B, 8260B
35	3	Methyl Chloride	601, 624, 8021B, 8260B
36	34418	Methylene Chloride	601, 624, 8021B, 8260B
37	34516	1,1,2,2-Tetrachloroethane	601, 624, 8021B, 8260B
38	34475	Tetrachloroethylene	601, 624, 8021B, 8260B
39	34010	Toluene	602, 624, 8021B, 8260B
40	34549	1,2-Trans-Dichloroethylene	601, 624, 8021B, 8260B
41	34506	1,1,1-Trichloroethane	601, 624, 8021B, 8260B
42	34511	1,1,2-Trichloroethane	601, 624, 8021B, 8260B
43	39180	Trichloroethylene	601, 624, 8021B, 8260B
44	39175	Vinyl Chloride	601, 624, 8021B, 8260B
		<b>Semi-Volatile Pollutants</b>	
45	34586	2-Chlorophenol	604, 625, 8041, 8270C
46	34601	2,4-Dichlorophenol	604, 625, 8041, 8270C
47	34606	2,4-Dimethylphenol	604, 625, 8041, 8270C
48	34452	2-Methyl-4,6-Dinitrophenol	604, 625, 8041, 8270C



**ATTACHMENT B-1**

#	Compound	Pollutant ID	Toxic Pollutants	EPA Analytical Method
49		34616	2,4-Dinitrophenol	604, 625, 8041, 8270C
50		34591	2-Nitrophenol	604, 625, 8041, 8270C
51		34646	4-Nitrophenol	604, 625, 8041, 8270C
52			3-Methyl-4-Chlorophenol	604, 625, 8041, 8270C
53		39032	Pentachlorophenol	604, 625, 8041, 8270C
54		34694	Phenol	604, 625, 8041, 8270C
55		34624	2,4,6-Trichlorophenol	604, 625, 8041, 8270C
56		34205	Acenaphthene	610, 625, 8100, 8270C
57		34200	Acenaphthylene	610, 625, 8100, 8270C
58		34220	Anthracene	610, 625, 8100, 8270C
59		39120	Benzidine	625, 8270C
60		34526	Benzo (a) Anthracene	610, 625, 8100, 8270C
61		34247	Benzo (a) Pyrene	610, 625, 8100, 8270C
62		34230	Benzo (b) Fluoranthene	610, 625, 8100, 8270C
63		34521	Benzo (g,h,i) Perylene	610, 625, 8100, 8270C
64		34242	Benzo (k) Fluoranthene	610, 625, 8100, 8270C
65		34278	Bis (2-Chloroethoxy) Methane	611, 625, 8270C
66		34283	Bis (2-Chloroisopropyl) Ether	611, 625, 8111, 8270C
67		34273	Bis (2-Chloroethyl) Ether	611, 625, 8111, 8270C
68		39100	Bis (2-Ethylhexyl) Phthalate	606, 625, 8061A, 8270C
69		34636	4-Bromophenyl Phenyl Ether	611, 625, 8111, 8270C
70		34292	Butylbenzyl Phthalate	606, 625, 8061A, 8270C
71		34581	2-Chloronaphthalene	612, 625, 8100, 8270C
72		34641	4-Chlorophenyl Phenyl Ether	611, 625, 8111, 8270C
73		34320	Chrysene	610, 625, 8100, 8270C
74		34556	Dibenzo (a,h) Anthracene	610, 625, 8100, 8270C
75		34536	1,2-Dichlorobenzene	601, 602, 612, 624, 625, 8021B, 8270C
76		34566	1,3-Dichlorobenzene	601, 602, 612, 624, 625, 8021B, 8270C
77		34571	1,4-Dichlorobenzene	601, 602, 612, 624, 625, 8021B, 8270C
78		34631	3,3-Dichlorobenzidine	625, 8270C
79		34336	Diethyl Phthalate	606, 625, 8061A, 8270C
80		34341	Dimethyl Phthalate	606, 625, 8061A, 8270C
81		34596	Di-n-Octyl Phthalate	606, 625, 8061A, 8270C
82		34611	2,4-Dinitrotoluene	609, 625, 8091, 8270C
83		34626	2,6-Dinitrotoluene	609, 625, 8091, 8270C
84		39110	Di-n-Butyl Phthalate	606, 625, 8061A, 8270C
85		34346	1,2-Diphenylhydrazine	625, 8270C
86		34376	Fluoranthene	610, 625, 8100, 8270C
87		34381	Fluorene	610, 625, 8100, 8270C
88		39700	Hexachlorobenzene	612, 625, 8120A, 8270C
89		39702	Hexachlorobutadiene	612, 625, 8120A, 8270C
90		34386	Hexachlorocyclopentadiene	612, 8120A, 8270C
91		34396	Hexachloroethane	616, 625, 8120A, 8270C

ATTACHMENT B-1

#	Compound	Pollutant ID	Toxic Pollutants	EPA Analytical Method
92		34403	Indeno (1,2,3-cd) Pyrene	610, 625, 8100, 8270C
93		34408	Isophorone	609, 625, 8270C
94		34696	Napthalene	610, 625, 8100, 8270C
95		34447	Nitrobenzene	609, 625, 8091, 8270C
96		34438	N-Nitrosodimethylamine	607, 625, 8070A, 8270C
97		34428	N-Nitrosodi-n-Propylamine	607, 625, 8070A, 8270C
98		34433	N-Nitrosodiphenylamine	607, 8070A, 8270C
99		34461	Phenanthrene	610, 625, 8100, 8270C
100		34469	Pyrene	610, 625, 8100, 8270C
101		34551	1,2,4-Trichlorobenzene	612, 625, 8120A, 8270C
			<b>Pesticides</b>	
102		39330	Aldrin	608, 8081A
103		39336	Alpha-BHC	608, 8081A
104		39338	beta-BHC	608, 8081A
105		39340	Gamma-BHC	608, 8081A
106		34198	delta-BHC	608, 8081A
107		39350	Chlordane	608, 8081A
108		39300	4,4'-DDT	608, 8081A
109		39320	4,4'-DDE	608, 8081A
110		39310	4,4'-DDD	608, 8081A
111		39380	Dieldrin	608, 8081A
112		78428	Alpha-Endosulfan	608, 8081A
113		34356	beta-Endosulfan	608, 8081A
114		34351	Endosulfan Sulfate	608, 8081A
115		39390	Endrin	608, 8081A
116		34366	Endrin Aldehyde	608, 8081A
117		39410	Heptachlor	608, 8081A
118		39420	Heptachlor Epoxide	608, 8081A
119-125		4166	PCBs)	608, 8082
126		39400	Toxaphene	608, 8081A
			<b>Miscellaneous receiving water Monitoring parameters</b>	
		4	pH of receiving water	
		2	Hardness (mg/L as CaCO3)	
			Salinity of receiving water (mg/L)	
			Receiving water flow rate (cfs)	