

CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD
LOS ANGELES REGION

MONITORING AND REPORTING PROGRAM NO. 8382
for
HOME DEPOT, U.S.A. (FORMER ITT AEROSPACE CONTROLS FACILITY)
(CA0064467)

I. Reporting Requirements

- A. Home Depot, Inc. (hereinafter Home Depot or Discharger) shall implement this monitoring program on the effective date of this Order. All monitoring reports shall be submitted quarterly and must be received by the Los Angeles Regional Water Quality Control Board (Regional Board) by the dates in the following schedule. All monitoring reports should be addressed to the Regional Board, Attention: Information Technology Unit. The first monitoring report under this Program is due by April 15, 2003.

<u>Reporting Period</u>	<u>Report Due</u>
January – March	April 15
April – June	July 15
July – September	October 15
October – December	January 15
Annual Summary Report	March 1

- B. If there is no discharge during any reporting period, the report shall so state.
- C. The Discharger shall submit an annual summary report, containing a discussion of the previous year's effluent monitoring data, as well as graphical and tabular summaries of the data. The data shall be submitted to the Regional Board on hard copy and on a 3 ½ " computer diskette. Submitted data must be IBM compatible, preferably using EXCEL software. In addition, the Discharger shall discuss the compliance record and the corrective actions taken or planned which may be needed to bring the discharge into full compliance with waste discharge requirements. This annual report is to be received by the Regional Board by March 1 of each year following the calendar year of data collection.
- D. The Discharger shall inform the Regional Board well in advance of any proposed construction activity that could potentially affect compliance with applicable requirements.

II. Effluent Monitoring Requirements

- A. A sampling station shall be established for each point of discharge and shall be located where representative samples of that effluent can be obtained.

- B. This Regional Board shall be notified in writing of any change in the sampling stations once established or in the methods for determining the quantities of pollutants in the individual waste streams.
- C. Pollutants shall be analyzed using the analytical methods described in 40 CFR 136.3, 136.4, and 136.5 (revised May 14, 1999); or, where no methods are specified for a given pollutant, by methods approved by this Regional Board or the State Board. Laboratories analyzing effluent samples and receiving water samples shall be certified by the California Department of Health Services and must include quality assurance/quality control (QA/QC) data in their reports.

The monitoring reports shall specify the analytical method used, the Method Detection Limit (MDL), and the Minimum Level (ML) for each pollutant. For the purpose of reporting compliance with numerical limitations, performance goals, and receiving water limitations, analytical data shall be reported by one of the following methods, as appropriate:

1. An actual numerical value for sample results greater than or equal to the ML; or,
2. "Detected, but Not Quantified (DNQ)" if results are greater than or equal to the laboratory's MDL but less than the ML; or,
3. "Not-Detected (ND)" for sample results less than the laboratory's MDL with the MDL indicated for the analytical method used.

Current MLs (Attachment T-1) are those published by the State Water Resources Control Board in the *Policy for the Implementation of Toxics Standards for Inland Surface Waters, Enclosed Bays, and Estuaries of California, March 2, 2000*.

- D. Where possible, the MLs employed for effluent analyses shall be lower than the permit limits established for a given parameter. If the ML value is not below the effluent limitation, then the lowest ML value and its associated analytical method shall be selected for compliance purposes. At least once a year, the Discharger shall submit a list of the analytical methods employed for each test and associated laboratory QA/QC procedures.

The Regional Board, in consultation with the State Board Quality Assurance Program, shall establish an ML that is not contained in Attachment T-1 to be included in the Discharger's permit in any of the following situations:

1. When the pollutant under consideration is not included in Attachment T-1;
2. When the Discharger and Regional Board agree to include in the permit a test method that is more sensitive than that specified in 40 CFR 136 (revised May 14, 1999);

3. When the Discharger agrees to use an ML that is lower than that listed in Attachment T-1;
 4. When the Discharger demonstrates that the calibration standard matrix is sufficiently different from that used to establish the ML in Attachment T-1, and proposes an appropriate ML for their matrix; or,
 5. When the Discharger uses a method whose quantification practices are not consistent with the definition of an ML. Examples of such methods are the USEPA-approved method 1613 for dioxins and furans, method 1624 for volatile organic substances, and method 1625 for semi-volatile organic substances. In such cases, the Discharger, the Regional Board, and the State Board shall agree on a lowest quantifiable limit and that limit will substitute for the ML for reporting and compliance determination purposes.
- E. Water/wastewater samples must be analyzed within allowable holding time limits as specified in 40 CFR Part 136.3. All QA/QC items must be run on the same dates the samples were actually analyzed, and the results shall be reported in the Regional Board format, when it becomes available, and submitted with the laboratory reports. Proper chain of custody procedures must be followed, and a copy of the chain of custody shall be submitted with the report.
- F. For parameters where both monthly average and daily maximum limits are specified but where the monitoring frequency is less than four times a month, the following procedure shall apply: If an analytical result is greater than the monthly average limit, the sampling frequency shall be increased (within one week of receiving the laboratory results) to a minimum of once weekly at equal intervals until at least four consecutive weekly samples have been obtained and compliance with the monthly average limit has been demonstrated again, and the Discharger has set forth for the approval of the Executive Officer, a program which ensures future compliance with the monthly average limit.

III. Effluent Monitoring Program

- A. The following shall constitute the effluent monitoring program for the final effluent:

Constituent	Units	Type of Sample	Sampling Frequency ^{1/}
Total waste flow ^{2/}	gal/day	----	daily
PH	Standard Units	grab	monthly ^{3/}
Turbidity	NTU	grab	monthly ^{3/}
Settleable solids	ml/L	grab	monthly ^{3/}
Total suspended solids	mg/L	grab	monthly ^{3/}
Oil and grease	mg/L	grab	monthly
Sulfate	mg/L	grab	monthly
Chloride	mg/L	grab	monthly
Sulfides	mg/L	grab	monthly
Nitrate-N + Nitrite-N	mg/L	grab	monthly

Constituent	Units	Type of Sample	Sampling Frequency ^{1/}
Ammonia	mg/L	grab	monthly
BOD ₅ 20°C	mg/L	grab	quarterly
Arsenic	µg/L	grab	monthly ^{3/}
Cadmium	µg/L	grab	monthly ^{3/}
Copper	µg/L	grab	monthly ^{3/}
Chromium III	µg/L	grab	monthly ^{3/}
Chromium VI	µg/L	grab	monthly ^{3/}
Lead	µg/L	grab	monthly ^{3/}
Mercury	µg/L	grab	monthly ^{3/}
Nickel	µg/L	grab	monthly ^{3/}
Selenium	µg/L	grab	monthly ^{3/}
Silver	µg/L	grab	monthly ^{3/}
Zinc	µg/L	grab	monthly ^{3/}
Trichloroethene	µg/L	grab	monthly ^{3/}
Tetrachloroethene	µg/L	grab	monthly ^{3/}
1,1,1-trichloroethane	µg/L	grab	monthly ^{3/}
1,1-Dichloroethene	µg/L	grab	monthly ^{3/}
Methyl tertiary butyl ether	µg/L	grab	monthly ^{3/}
Total Petroleum Hydrocarbons (both gasoline and diesel fractions) ^{4/}	µg/L	grab	monthly
Phenols	mg/L	grab	quarterly ^{5/}
Phenolic compounds (chlorinated)	µg/L	grab	quarterly ^{5/}
Perchlorate	µg/L	grab	quarterly ^{5/}
Acetone	µg/L	grab	quarterly ^{5/}
Benzene	µg/L	grab	quarterly ^{5/}
Carbon tetrachloride	µg/L	grab	quarterly ^{5/}
Ethyl benzene	µg/L	grab	quarterly ^{5/}
Tertiary butyl alcohol	µg/L	grab	quarterly ^{5/}
Toluene	µg/L	grab	quarterly ^{5/}
Xylene	µg/L	grab	quarterly ^{5/}
Vinyl chloride	µg/L	grab	quarterly ^{5/}
1,4-dichlorobenzene	µg/L	grab	quarterly ^{5/}
1,2-dichloroethene	µg/L	grab	quarterly ^{5/}
Remaining priority pollutants (see page T-15)	µg/L	grab	semi-annually
Toxicity – acute	% survival	grab	semi-annually ^{6/}
Toxicity – Chronic	TU _c	grab	semi-annually

1/ If any constituent exceeds the limit in Order No. R4-2002-0185, the discharge shall be terminated and shall only be resumed after remedial measures have been implemented, and full compliance with the requirements has been ascertained.

- 2/ Actual monitored flow from the outfall (not the maximum permitted flow) shall be reported.
- 3/ Shall be monitored weekly for the first three months. If all the analytical results are below the discharge limits for the first three month period, monitoring thereafter shall revert to the frequency specified in the monitoring period. If any of the analytical results exceed the requirement in the first three month period, then the monitoring shall be continued on a weekly basis till three month continuous compliance of the limit is achieved.
- 4/ Analyses using USEPA Methods 418.1 and 8015 (Modified).
- 5/ Shall be monitored monthly for the first six months. If all the analytical results are below the discharge limits for the first six months period, monitoring thereafter shall revert to the frequency specified in the monitoring period. If any of the analytical results exceed the requirement in the first six month period, then the monitoring shall be continued on a monthly basis till three month continuous compliance of the limit is achieved.
- 6/ Monitoring shall be on a quarterly basis for first two years, from the date of adoption of the permit. Monitoring thereafter shall revert to the frequency specified in the monitoring program.

IV. Toxicity Monitoring Requirements

A. Acute Toxicity Monitoring Program

1. The Discharger shall conduct acute toxicity tests on effluent grab samples by methods specified in 40 CFR Part 136 which cites USEPA's *Methods for Measuring the Acute Toxicity of Effluents to Freshwater and Marine Organisms*, August, 1991 (EPA/600/4-90/027) or a more recent edition to ensure compliance in 100 % effluent.
2. The fathead minnow, *Pimephales promelas*, shall be used as the test species for fresh water discharges and the topsmelt, *Atherinops affinis*, shall be used as the test species for brackish effluent. The method for topsmelt is found in USEPA's *Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to West Coast Marine and Estuarine to Freshwater Organisms*, First Edition, August, 1995 (EPA/600/4-95/136).
3. In lieu of conducting the standard acute toxicity testing with the fathead minnow, the Discharger may elect to report the results or endpoint from the first 48 hours of the chronic toxicity test as the results of the acute toxicity test.

B. Chronic Toxicity Effluent Monitoring Program:

1. The Discharger shall conduct critical life stage chronic toxicity tests on effluent samples (24-hour composite) or receiving water samples in accordance with EPA's Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms, Third Edition, July 1994 (EPA/600/4-91/002) or EPA's Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms, August 1995, (EPA/600/R-95/136).

2. Effluent samples shall be collected after all treatment processes and before discharge to the receiving water.
 3. Test Species and Methods:
 - a. The Discharger shall conduct tests as follows: with a vertebrate, an invertebrate, and an alga for the first three suites of tests. After the screening period, monitoring shall be conducted using the most sensitive species.
 - b. Re-screening is required every 15 months. The Discharger shall re-screen with the three species listed above and continue to monitor with the most sensitive species. If the first suite of re-screening tests demonstrates that the same species is the most sensitive than the re-screening does not need to include more than one suite of tests. If a different species is the most sensitive or if there is ambiguity then the Discharger shall proceed with suites of screening tests for a minimum of three, but not to exceed five suites.
 - c. The presence of chronic toxicity shall be estimated as specified using West Coast marine organisms according to EPA's Short-Term Methods for Estimating Chronic Toxicity of Effluent and Receiving Waters to West Coast Marine and Estuarine Organisms, August, 1995 (EPA/600/R-95/136).
- C. Quality Assurance
1. Concurrent testing with a reference toxicant shall be conducted. Reference toxicant tests shall be conducted using the same test conditions as the effluent toxicity tests (e.g., same test duration, etc).
 2. If either the reference toxicant test or effluent test does not meet all test acceptability criteria (TAC) as specified in the test methods manuals (EPA/600/4-91/002, EPA/600/R-95/136, or EPA/600/4-90/027F), then the Discharger must re-sample and re-test within 14 days of notification by the laboratory of an invalid test.
 3. Control and dilution water shall be receiving water or laboratory water as described in the manual. If the dilution water used is different from the culture water, a second control using culture water shall be used.
- D. Accelerated Monitoring
1. If toxicity exceeds the limitations (as defined in Order No. R4-2002-0185, Section I.B.3.b.i), then the Discharger shall immediately implement accelerated testing, as specified at Section I.B.3.b.ii. The Discharger shall ensure that they receive results of a failing toxicity test within 24 hours of the completion of the test and the additional tests shall begin within 3 business days of receipt of the

results. If the accelerated testing shows consistent toxicity, the Discharger shall immediately implement the Initial Investigation of the TRE Workplan.

2. If implementation of the initial investigation TRE workplan indicates the source of toxicity (e.g., a temporary plant upset, etc.), then the Discharger may discontinue the TIE.
 3. The first step in the initial Investigation TRE Workplan for downstream receiving water toxicity can be a toxicity test protocol designed to determine if the effluent from Outfall 001 causes or contributes to the measured downstream chronic toxicity. If this first step TRE testing shows that the Outfall 001 effluent does not cause or contribute to downstream chronic toxicity, using EPA's Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms, August 1995, (EPA/600/R-95/136). Then a report on this testing shall be submitted to the Board and the TRE will be considered to be completed. Routine testing in accordance with MRP No. 7697 shall be continued thereafter.
- E. Steps in Toxicity Reduction Evaluation (TRE) and Toxicity Identification Evaluation (TIE)
1. Following a TRE trigger, the Discharger shall initiate a TRE in accordance with the facility's initial investigation TRE workplan. At a minimum, the Discharger shall use EPA manuals EPA/600/2-88/070 (industrial) or EPA/833B-99/002 (municipal) as guidance. The Discharger shall expeditiously develop a more detailed TRE workplan for submittal to the Executive Officer within 30 days of the trigger, which will include, but not be limited to:
 - a. Further actions to investigate and identify the cause of toxicity;
 - b. Actions the Discharger will take to mitigate the impact of the discharge and prevent the recurrence of toxicity;
 - c. Standards the Discharger will apply to consider the TRE complete and to return to normal sampling frequency; and,
 - d. A schedule for these actions
 2. The following is a stepwise approach in conducting the TRE:
 - a. Step 1 - Basic data collection. Data collected for the accelerated monitoring requirements may be used to conduct the TRE;
 - b. Step 2 - Evaluates optimization of the treatment system operation, facility housekeeping, and the selection and use of in-plant process chemicals;
 - c. If Steps 1 and 2 are unsuccessful, Step 3 implements a TIE and employment of all reasonable efforts and using currently available TIE methodologies. The objective of the TIE is to identify the substance or

combination of substances causing the observed toxicity;

- d. Assuming successful identification or characterization of the toxicant(s), Step 4 evaluates final effluent treatment options;
- e. Step 5 evaluates in-plant treatment options; and,
- f. Step 6 consists of confirmation once a toxicity control method has been implemented.

Many recommended TRE elements parallel source control, pollution prevention, and storm water control program best management practices (BMPs). To prevent duplication of efforts, evidence of implementation of these control measures may be sufficient to comply with TRE requirements. By requiring the first steps of a TRE to be accelerated testing and review of the facility's TRE workplan, a TRE may be ended in its early stages. All reasonable steps shall be taken to reduce toxicity to the required level. The TRE may be ended at any stage if monitoring indicates there is no longer toxicity (or six consecutive chronic toxicity results are less than or equal to 1.0 TU_c).

3. The Discharger may initiate a TIE as part of the TRE process to identify the cause(s) of toxicity. The Discharger shall use the EPA acute and chronic manuals, EPA/600/6-91/005F (Phase I)/EPA/600/R-96-054 (for marine), EPA/600/R-92/080 (Phase II), and EPA-600/R-92/081 (Phase III) as guidance.
4. If a TRE/TIE is initiated prior to completion of the accelerated testing schedule required by Part I.B.4.a.2 and Part I.B.4.b.2 of this permit, then the accelerated testing schedule may be terminated, or used as necessary in performing the TRE/TIE, as determined by the Executive Officer.
5. Toxicity tests conducted as part of a TRE/TIE may also be used for compliance, if appropriate.
6. The Board recognizes that toxicity may be episodic and identification of causes of and reduction of sources of toxicity may not be successful in all cases. Consideration of enforcement action by the Board will be based in part on the Discharger's actions and efforts to identify and control or reduce sources of consistent toxicity.

F. Reporting

1. The Discharger shall submit a full report of the toxicity test results, including any accelerated testing conducted during the month as required by this permit. Test results shall be reported in Toxicity Units (percent survival or TU_c) with the discharge monitoring reports (DMR) for the month in which the test is conducted.

If an initial investigation indicates the source of toxicity and accelerated testing is unnecessary, pursuant to Section IV.D.1., those results shall also be submitted

with the DMR for the period in which the Investigation occurred.

2. The full report shall be submitted on or before the end of the month in which the DMR is submitted.
3. The full report shall consist of (1) the results; (2) the dates of sample collection, initiation, and completion of each toxicity tests; (3) the acute toxicity limit or chronic toxicity limit or trigger as described in 4.A.1 and 4.B.1.
4. Test results for toxicity tests also shall be reported according to the appropriate manual chapter on Report Preparation and shall be attached to the DMR. Routine reporting shall include, at a minimum, as applicable, for each test:
 5. sample date(s);
 6. test initiation date;
 7. test species;
 8. end point values for each dilution (e.g., number of young, growth rate, percent survival);
 9. NOEC value(s) in percent effluent;
 10. IC₁₅, IC₂₅, IC₄₀ and IC₅₀ values in percent effluent;
 11. TU_c values $\left(TU_c = \frac{100}{NOEC} \right)$;
 12. Mean percent mortality (\pm standard deviation) after 96 hours in 100% effluent (if applicable);
 13. NOEC and LOEC values for reference toxicant test(s);
 14. IC₂₅ value for reference toxicant test(s);
 15. Any applicable control charts; and
 16. Available water quality measurements for each test (e.g., pH, D.O., temperature, conductivity, hardness, salinity, ammonia).
17. The Discharger shall provide a compliance summary, which includes a summary table of toxicity data from at least eleven of the most recent samples.

The Discharger shall notify, by telephone or electronically, this Regional Board of any toxicity exceedance of the limit or trigger within 24 hours of receipt of the results followed by a written report within 14 calendar days of receipt of the results. The verbal or electronic notification shall include the exceedance and

the plan the Discharger will pursue. The written report shall describe actions the Discharger has taken or will take to investigate and correct the cause(s) of toxicity. It may also include a status report on any actions required by the permit, with a schedule for actions not yet completed. If no actions have been taken, the reasons shall be given.

V. Interim Monitoring and Reporting

Pursuant to the California Water Code, Section 13267, the Discharger is required to submit data sufficient for: (1) determining if water quality-based effluent limitations for priority pollutants are required, and (2) to calculate effluent limitations, if required. The *Policy for the Implementation of Toxics Standards for Inland Surface Waters, Enclosed Bays, and Estuaries of California* (March 2, 2000) requires that the data be provided. Therefore, the Discharger shall conduct the following interim monitoring program for all California Toxics Rule priority pollutants until December 2004, or until ordered otherwise by the Regional Board. As described in Section I.A of this Program, monitoring reports must be submitted quarterly. The Discharger shall ensure that at least four samples are collected in the interim monitoring period (two per year), the results of which will be submitted along with the corresponding quarterly reports.

- A. Effluent monitoring shall be conducted for all pollutants at the NPDES discharge point during periods when a discharge occurs. The data that is collected as part of the regular monitoring program in III.A can be substituted for interim effluent monitoring requirement.
- B. Receiving water monitoring station shall be at 50 feet upstream from the discharge point of storm drain to Los Angeles River.
- C. Monitoring frequency and type of sample of the effluent and the receiving water shall be collected and analyzed for all toxic pollutants listed below: Effluent and receiving water data shall be collected on the same day.

Constituent	Units	Type of Sample	Monitoring Frequency
pH	Standard Units	grab	semiannually
Hardness (as CaCO ₃)	mg/L	grab	semiannually
PAHs	µg/L	grab	semiannually
Arsenic	µg/L	grab	semiannually
Antimony	µg/L	grab	semiannually
Cadmium	µg/L	grab	semiannually
Copper	µg/L	grab	semiannually
Chromium III	µg/L	grab	semiannually
Chromium VI	µg/L	grab	semiannually
Lead	µg/L	grab	semiannually
Mercury	µg/L	grab	semiannually
Nickel	µg/L	grab	semiannually
Selenium	µg/L	grab	semiannually

Constituent	Units	Type of Sample	Monitoring Frequency
Silver	µg/L	grab	semiannually
Thallium	µg/L	grab	semiannually
Perchlorate	µg/L	grab	semiannually
Cyanide	µg/L	grab	semiannually
Acenaphthene	µg/L	grab	semiannually
Anthracene	µg/L	grab	semiannually
Benzo (a) Anthracene	µg/L	grab	semiannually
Benzo (a) Pyrene	µg/L	grab	semiannually
Benzo (b) Fluoranthene	µg/L	grab	semiannually
Benzo (k) Fluoranthene	µg/L	grab	semiannually
Chrysene	µg/L	grab	semiannually
Dibenzo (a,h) Anthracene	µg/L	grab	Semiannually
Fluoranthene	µg/L	grab	semiannually
Fluorene	µg/L	grab	semiannually
Indeno (1,2,3-cd) Pyrene	µg/L	grab	semiannually
Pyrene	µg/L	grab	semiannually
Aldrin	µg/L	grab	semiannually
Alpha-BHC	µg/L	grab	semiannually
Beta-BHC	µg/L	grab	semiannually
Chlordane	µg/L	grab	semiannually
Dieldrin	µg/L	grab	semiannually
Alpha-Endosulfan	µg/L	grab	semiannually
Beta-Endosulfan	µg/L	grab	semiannually
Heptachlor	µg/L	grab	semiannually
Heptachlor Epoxide	µg/L	grab	semiannually
4,4-DDT	µg/L	grab	semiannually
4,4-DDE	µg/L	grab	semiannually
4,4-DDD	µg/L	grab	Semiannually
Arochlor 1242	µg/L	grab	semiannually
Arochlor 1254	µg/L	grab	semiannually
Arochlor 1221	µg/L	grab	semiannually
Arochlor 1232	µg/L	grab	semiannually
Arochlor 1248	µg/L	grab	semiannually
Arochlor 1260	µg/L	grab	semiannually
Arochlor 1016	µg/L	grab	semiannually
Toxaphene	µg/L	grab	semiannually
Beryllium	µg/L	grab	semiannually
Asbestos	Fibers/L	grab	semiannually
Acrolein	µg/L	grab	semiannually
Acrylonitrile	µg/L	grab	semiannually
Benzene	µg/L	grab	semiannually
Bromoform	µg/L	grab	Semiannually

Constituent	Units	Type of Sample	Monitoring Frequency
Carbon tetrachloride	µg/L	grab	semiannually
Chlorobenzene	µg/L	grab	semiannually
Chlorodibromomethane	µg/L	grab	semiannually
Chloroethane	µg/L	grab	semiannually
2-Chloroethylvinyl ether	µg/L	grab	semiannually
Chloroform	µg/L	grab	semiannually
Dichlorobromomethane	µg/L	grab	semiannually
1,1-Dichloroethane	µg/L	grab	semiannually
1,2-Dichloroethane	µg/L	grab	semiannually
1,1-Dichloroethylene	µg/L	grab	semiannually
1,2-Dichloropropane	µg/L	grab	semiannually
1,3-Dichloropropylene	µg/L	grab	semiannually
Ethylbenzene	µg/L	grab	semiannually
Methyl bromide	µg/L	grab	Semiannually
Methyl chloride	µg/L	grab	semiannually
Methylene chloride	µg/L	grab	semiannually
1,1,2,2-Tetrachloroethane	µg/L	grab	semiannually
Tetrachloroethylene	µg/L	grab	semiannually
Toluene	µg/L	grab	semiannually
1,2-Trans-dichloroethylene	µg/L	grab	semiannually
1,1,1-Trichloroethane	µg/L	grab	semiannually
1,1,2-Trichloroethane	µg/L	grab	semiannually
Trichloroethylene	µg/L	grab	semiannually
Vinyl chloride	µg/L	grab	semiannually
2-Chlorophenol	µg/L	grab	semiannually
2,4-Dichlorophenol	µg/L	grab	semiannually
2,4-Dimethylphenol	µg/L	grab	semiannually
2-Methyl-4,6-Dinitrophenol	µg/L	grab	Semiannually
2,4-Dinitrophenol	µg/L	grab	semiannually
2-Nitrophenol	µg/L	grab	semiannually
4-Nitrophenol	µg/L	grab	semiannually
3-Methyl-4-Chlorophenol	µg/L	grab	semiannually
Pentachlorophenol	µg/L	grab	semiannually
2,4,6-Trichlorophenol	µg/L	grab	semiannually
Acenaphthylene	µg/L	grab	semiannually
Benzidine	µg/L	grab	semiannually
Benzo (g,h,i) Perylene	µg/L	grab	semiannually
Bis (2-Chloroethoxy) Methane	µg/L	grab	semiannually
Bis (2-Chloroethyl) Ether	µg/L	grab	semiannually
Bis (2-Chloroisopropyl) Ether	µg/L	grab	semiannually
Bis (2-Ethylhexyl) Phthalate	µg/L	grab	semiannually
4-Bromophenyl Phenyl Ether	µg/L	grab	Semiannually
Butylbenzyl Phthalate	µg/L	grab	semiannually
2-Chloronaphthalene	µg/L	grab	semiannually

Constituent	Units	Type of Sample	Monitoring Frequency
4-Chlorophenyl Phenyl Ether	µg/L	grab	Semiannually
1,2-Dichlorobenzene	µg/L	grab	semiannually
1,3-Dichlorobenzene	µg/L	grab	semiannually
1,4-Dichlorobenzene	µg/L	grab	semiannually
3,3-Dichlorobenzidine	µg/L	grab	semiannually
Diethyl Phthalate	µg/L	grab	semiannually
Dimethyl Phthalate	µg/L	grab	semiannually
Di-n-Butyl Phthalate	µg/L	grab	semiannually
2,4-Dinitrotoluene	µg/L	grab	semiannually
2,6-Dinitrotoluene	µg/L	grab	semiannually
Di-n-Octyl Phthalate	µg/L	grab	semiannually
1,2-Diphenylhydrazine	µg/L	grab	semiannually
Hexachlorobenzene	µg/L	grab	semiannually
Hexachlorobutadiene	µg/L	grab	semiannually
Hexachlorocyclopentadiene	µg/L	grab	semiannually
Hexachloroethane	µg/L	grab	semiannually
Isophorone	µg/L	grab	semiannually
Napthalene	µg/L	grab	semiannually
Nitrobenzene	µg/L	grab	Semiannually
N-Nitrosodimethylamine	µg/L	grab	semiannually
N-Nitrosodi-n-Propylamine	µg/L	grab	semiannually
N-Nitrosodiphenylamine	µg/L	grab	semiannually
Phenanthrene	µg/L	grab	semiannually
1,2,4-Trichlorobenzene	µg/L	grab	semiannually
Gamma-BHC	µg/L	grab	semiannually
Delta-BHC	µg/L	grab	Semiannually
Endosulfan Sulfate	µg/L	grab	semiannually
Endrin	µg/L	grab	semiannually
Endrin Aldehyde	µg/L	grab	Semiannually

- D. Monitoring for TCDD Equivalents –The Discharger shall conduct effluent/receiving water monitoring for the presence of the 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD or Dioxin) congeners. The monitoring shall be a grab sample with a minimum frequency of once during dry weather and once during wet weather for 1 year. The Discharger shall calculate Toxic Equivalence (TEQ) for each congener by multiplying its analytical concentration by the appropriate Toxicity Equivalence Factors (TEF). Compliance with the dioxin limitation shall be determined by the summation of the 17 individual TEQs.

Congeners	TEF
2,3,7,8-tetra CDD	1.0
1,2,3,7,8-penta CDD	1.0
1,2,3,4,7,8-hexa CDD	0.1
1,2,3,6,7,8-hexa CDD	0.1
1,2,3,7,8,9-hexa CDD	0.1
1,2,3,4,6,7,8-hepta CDD	0.01
Octa CDD	0.0001
2,3,7,8-tetra CDF	0.1
1,2,3,7,8-penta CDF	0.05
2,3,4,7,8-penta CDF	0.5
1,2,3,4,7,8-hexa CDF	0.1
1,2,3,6,7,8-hexa CDF	0.1
1,2,3,7,8,9-hexa CDF	0.1
2,3,4,6,7,8-hexa CDF	0.1
1,2,3,4,6,7,8-hepta CDF	0.01
1,2,3,4,7,8,9-hepta CDF	0.01
Octa CDF	0.0001

Ordered by: _____
Dennis A. Dickerson
Executive Officer

Date: December 12, 2002